

Abstract

This dissertation is focused on 30 kDa protein phosphodiesterase (Pde) and on the regulation of its function through the interaction with 28 kDa adaptor protein 14-3-3. These two proteins participate in G-protein signal transduction pathways, especially in the process of light signal transduction. It is assumed that Pde binds to the $G_i\beta\gamma$ complex of G-protein called transducin and through this interaction it inhibits the reassociation of $G_i\beta\gamma$ with $G_i\alpha$ thus reducing the visual signal transfer. This process is thought to participate in a long-term light adaptation. The regulation of Pde function is further regulated by its phosphorylation and subsequent binding to the 14-3-3 protein. It has been speculated that the 14-3-3 binding plays a key role in the inhibition of the interaction between phosphorylated Pde (Pde-PP) and $G_i\beta\gamma$. The formation of the 14-3-3/Pde-PP complex leads to the reassociation of $G_i\beta\gamma$ with $G_i\alpha$ and consequently to the amplification of visual signal transfer. Nevertheless, the mechanism by which the 14-3-3 protein binding inhibits the interaction between Pde and $G_i\beta\gamma$ remains elusive.

The main aims of this dissertation were: (i) to investigate the structure of Pde in its apo-state (in the absence of the binding partner) and in the complex with 14-3-3, and (ii) to suggest the mechanism of the 14-3-3-mediated regulation of Pde function. The structure of Pde and the 14-3-3/Pde-PP complex was studied using various biophysical methods including small-angle X-ray scattering (SAXS), NMR, H/D exchange coupled to mass spectrometry (HDX-MS) and fluorescence techniques. Our data suggested that the N-terminal domain of Pde (Pde-ND) is intrinsically disordered protein. The phosphorylation of Pde affects conformation of both its domains - unstructured Pde-ND as well as structured C-terminal domain (Pde-CD). The 14-3-3 protein binds Pde-PP with the binding affinity in micromolar range and the complex formation affects the conformation of Pde especially within Pde-ND. Both phosphorylation sites are essential for the complex formation. The majority of Pde-ND, which accounts for the most of the $G_i\beta$ binding surface, is located either in the central channel of the 14-3-3 dimer or in the close vicinity of the 14-3-3 outer surface. Therefore, our data suggest that the 14-3-3 binding masks the majority of the $G_i\beta$ binding surface of Pde and thus inhibits its interaction with $G_i\beta$. In addition, NMR and HDX-MS measurements revealed that Pde-PP remains highly flexible after the 14-3-3 binding, which indicates the formation of a “fuzzy” complex. Our data also showed that the 14-3-3 binding slows down dephosphorylation of Pde-PP *in vitro*.