Přílohy

Příloha č. 1 *Protokol pro izolaci DNA-Genomic DNA Mini Kit, převzato z www.geneaid.com*

Genomic DNA Mini Kit (Tissue)

For research use only

Sample: up to 30 mg of tissue (tailsnips, liver, kidney, brain, adipose tissue, earpunches, insects etc.)
Yield: 10-20 µg (0.5 cm of mouse tail, 20 mg of mouse liver), 20-50 µg (20 mg of mouse kidney)
Format: spin column
Time: within 30 minutes
Elution volume: 30-200 µl
Storage: dry at room temperature (15-25°C)

ISO 9001:2008 QMS

Introduction

The Genomic DNA Mini Kit (Tissue) was designed specifically for purifying total DNA (including genomic, mitochondrial and viral DNA) from a variety of tissue and insect samples. The provided micropestle can efficiently homogenize tissue samples to shorten the time in the Lysis Step. Proteinase K and chaotropic salt are used to lyse cells and degrade protein, allowing DNA to be easily bound by the glass fiber matrix of the spin column. Once any contaminants have been removed, using a Wash Buffer (containing ethanol), the purified DNA is eluted by a low salt Elution Buffer, TE or water. The entire procedure can be completed without phenol/chloroform extraction or alcohol precipitation. The purified DNA (approximately 20-30 kb) is suitable for use in PCR or other enzymatic reactions.

Quality Control

The quality of the Genomic DNA Mini Kit (Tissue) is tested on a lot-to-lot basis by isolating genomic DNA from a 20 mg mouse liver sample. The purified DNA (more than 10 µg with an A260/A280 ratio of 1.8-2.0) is quantified with a spectrophotometer and analyzed by electrophoresis.

Kit Contents

Component	GT004	GT050	GT100	GT300
GT Buffer	3 ml	30 ml	30 ml	75 ml
GBT Buffer	4 ml	40 ml	40 ml	75 ml
W1 Buffer	2 ml	45 ml	45 ml	130 ml
Wash Buffer* (Add Ethanol)	1 ml (4 ml)	25 ml (100 ml)	25 ml (100 ml)	50 ml (200 ml)
Proteinase K** (Add ddH_2O)	1 mg (0.1 ml)	11 mg (1.1 ml)	11 mg x 2 (1.1 ml x 2)	65 mg (6.5 ml)
Elution Buffer	1 ml	30 ml	30 ml	75 ml
GS Columns	4	50	100	300
2 ml Collection Tubes	8	100	200	600
Micropestle	**4**	**50**	**100**	**300**

Order Information

Product	Package Size	Catalogue Number
Genomic DNA Mini Kit (Blood/Cultured Cell)	**100/300 preps**	**GB100/300**
Genomic DNA Maxi Kit (Blood/Cultured Cell)	10/25 preps	GDM010/25
Genomic DNA Mini Kit (Tissue)	**50/100/300 preps**	**GT050/100/300**
gSYNC™ DNA Extraction Kit	50/100/300 preps	GS050/100/300
Genomic DNA Mini Kit (Plant)	**100 preps**	**GP100**
Genomic DNA Maxi Kit (Plant)	10/25 preps	GPM010/25
GENEzol™ DNA Reagent Plant	**100/200 rxns**	**GR100/200**
Presto™ Mini gDNA Yeast Kit	100/300 preps	GBY100/300
Presto™ Mini gDNA Bacteria Kit	**100/300 preps**	**GBB100/101/300/301**
Geneius™ Micro DNA Extraction Kit	100/300 preps	GMB100/300
Presto™ Buccal Swab gDNA Extrac on Kit	**100/300 preps**	**GSK100/300**
Presto™ 96 Well Blood gDNA Extraction Kit	4/10 x 96 preps	96GBP04/10
Presto™ 96 Well Plant gDNA Extraction Kit	**4/10 x 96 preps**	**96GPP04/10**

*Add absolute ethanol (see the bottle label for volume) to the Wash Buffer prior to initial use
**Add ddH_2O (see the bottle label for volume) to prepare Proteinase K (vortex to dissolve and spin down) and store at 4°C

Caution

GBT Buffer contains guanidine hydrochloride. During operation, always wear a lab coat, disposable gloves, and protective goggles.

Genomic DNA Mini Kit (Tissue) Functional Test Data

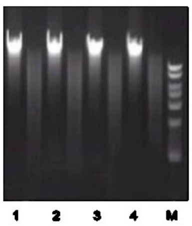


Figure 1. Genomic DNA from a variety of tissue samples was extracted using the Genomic DNA Mini Kit (Tissue). The purified genomic DNA (30-40 kb) was *EcoRI* digested and analyzed by electrophoresis on a 1% agarose gel.
1 = Mouse Liver
2 = Mouse Tail
3 = Fish Muscle
4 = Fruit Fly (Drosophila)
M = Geneaid 1 Kb DNA Ladder

Genomic DNA Mini Kit (Tissue) Protocol

IMPORTANT BEFORE USE

- Add ddH_2O (see the bottle label for volume) to prepare Proteinase K (vortex to dissolve and spin down) and store at 4ºC
- Add absolute ethanol (see the bottle label for volume) to the Wash Buffer prior to initial use
- Additional requirements: microcentrifuge tubes, absolute ethanol, (optional) RNase A (10 mg/ml), ddH_2O

Tissue Dissociation	• Cut up to **30 mg of animal tissue (or 0.5 cm of mouse tail)** then transfer it to a 1.5 ml microcentrifuge tube. NOTE: If tissue has a higher number of cells (e.g. spleen or liver), reduce the starting material to 10 mg. • Use the provided **Micropestle** to grind the tissue to a pulp. • Add **200 µl of GT Buffer** to the tube and homogenize the sample tissue by grinding. • Add **20 µl of Proteinase K** to the sample mixture then shake vigorously and incubate at 60ºC for 30 minutes. NOTE: During incubation, invert the tube every 5 minutes.
Step 1 Lysis	• Add **200 µl of GBT Buffer** then shake vigorously for 5 seconds. • **Incubate at 60ºC for at least 20 minutes to ensure the lysate is clear.** inkubace na 56ºC přes celou noc NOTE: During incubation, invert the tube every 5 minutes. If insoluble material is present following incubation, centrifuge for 2 minutes at 14-16,000 x g then transfer the supernatant to a new 1.5 ml microcentrifuge tube. At this time, preheat the required **Elution Buffer** (200 µl per sample) to 60ºC (for Step 4 DNA Elution). **Optional Step: RNA Degradation** (If RNA free gDNA is required, perform this optional step) • Following 60ºC incubation, add 4 µl of RNase A (10 mg/ml) to the sample lysate then shake vigorously. • Incubate at room temperature for 5 minutes.
Step 2 DNA Binding	• Add **200 µl of absolute ethanol** to the lysate then immediately shake vigorously for 10 seconds. NOTE: If precipitate appears, break it up as much as possible with a pipette. • Place a **GS Column** in a **2 ml Collection Tube**. • Transfer the mixture (including any precipitate) to the **GS Column** then centrifuge at 14-16,000 x g for 2 minutes. • Discard the **2 ml Collection Tube** then transfer the **GS Column** to a new **2 ml Collection Tube**.
Step 3 Wash	• **Add 400 µl of W1 Buffer to the GS Column then centrifuge at 14-16,000 x g for 30 seconds.** • **Discard the flow-through then place the GS Column back in the 2 ml Collection Tube.** • **Add 600 µl of Wash Buffer (make sure ethanol was added) to the GS Column.** • **Centrifuge at 14-16,000 x g for 30 seconds.** • **Discard the flow-through then place the GS Column back in the 2 ml Collection Tube.** • **Centrifuge for 3 minutes at 14-16,000 x g to dry the column matrix.**
Step 4 DNA Elution	Standard elution volume is 100 µl. If less sample is to be used, reduce the elution volume (30-50 µl) to increase DNA concentration. If higher DNA yield is required, repeat the DNA Elution step to increase DNA recovery and the total elution volume to approx. 200 µl. • Transfer the dried **GS Column** to a clean 1.5 ml microcentrifuge tube. přidáno 75µl Elution Buffer, • Add **100 µl of pre-heated Elution Buffer** or TE to the CENTER of the column matrix. 5 min absorpce, centrifuga • Let stand for at least 5 minutes to ensure the **Elution Buffer** or TE is completely absorbed. 2x opakováno • Centrifuge at 14-16,000 x g for 30 seconds to elute the purified DNA.

Troubleshooting

Problem	Possible Reasons/Solution
Clogged Column	**Too much tissue was used** • If using more than 30 mg of tissue, separate into multiple tubes. **Sample tissue was not lysed completely** • **Add additional Proteinase K and extend the incubation time in the Lysis Step.** • **Following the Lysis Step, centrifuge for 2 minutes at 14-16,000 x g to remove sample debris. Transfer the supernatant to a new microcentrifuge tube and proceed with the DNA Binding Step.** **Precipitate was formed at DNA Binding step** • Reduce the sample material. • Following ethanol addition, break up any precipitate as much as possible prior to loading GS Column.
Low Yield	**Sample tissue was not lysed completely** • Add additional Proteinase K and extend the incubation time in the Lysis Step. **Column was clogged at DNA Binding step** • Following the Lysis Step, remove the insoluble debris by centrifugation. • Prior to loading the column, break up the precipitate in the ethanol-added lysate. **Incorrect DNA Elution Step** • Ensure that the Elution Buffer or TE is added to the center of the GS Column matrix and is absorbed completely. **Incomplete DNA elution** • Elute twice to increase the DNA recovery.
Eluted DNA does not perform well in downstream applications	**Residual ethanol contamination** • Following the Wash Step, dry the GS Column by centrifuge at 14-16,000 x g or incubate at 60ºC for 5 minutes. **RNA/Protein contamination** • Perform optional RNA Degradation step/reduce the sample amount. **Genomic DNA was degraded** • **Use fresh samples or freeze fresh samples in liquid nitrogen immediately and store at -80ºC.**

Příloha č. 2 *Frekvence alel v jednotlivých populacích vygenerované programem FSTAT.*

Populace:	Zalomka	Cernica	Vrijeka	Mušnica	Ljuta
Locus: **BL130**					
N	28	30	25	8	4
p: 79	0.393	0.017	0.000	0.000	0.000
p: 85	0.107	0.100	0.900	0.188	1.000
p: 87	0.464	0.800	0.080	0.625	0.000
p: 89	0.018	0.033	0.020	0.188	0.000
p: 93	0.018	0.050	0.000	0.000	0.000
Locus: **BL144**					
N	28	28	25	7	4
p: 122	1.000	0.696	0.880	1.000	1.000
p: 126	0.000	0.054	0.000	0.000	0.000
p: 130	0.000	0.250	0.120	0.000	0.000
Locus: **Lsou05**					
N	28	29	24	9	4
p: 177	0.000	0.000	0.042	0.000	0.000
p: 185	0.536	0.000	0.083	0.000	0.000
p: 187	0.036	0.000	0.583	0.278	0.000
p: 189	0.000	0.000	0.063	0.167	0.000
p: 191	0.321	0.983	0.104	0.278	0.000
p: 193	0.000	0.017	0.063	0.000	0.125
p: 195	0.000	0.000	0.000	0.000	0.625
p: 201	0.036	0.000	0.063	0.000	0.000
p: 203	0.036	0.000	0.000	0.111	0.250
p: 205	0.036	0.000	0.000	0.056	0.000
p: 207	0.000	0.000	0.000	0.111	0.000
Locus: **BL161**					
N	30	30	26	9	4
p: 195	0.000	0.017	0.000	0.000	0.000
p: 197	0.983	0.983	1.000	1.000	1.000
p: 199	0.017	0.000	0.000	0.000	0.000
Locus: **BL1153**					
N	29	28	26	9	4
p: 220	0.741	0.857	1.000	0.833	1.000
p: 222	0.259	0.143	0.000	0.167	0.000
Locus: **Lsou19**					
N	30	30	26	9	4
p: 178	0.683	0.833	1.000	0.944	1.000
p: 180	0.317	0.017	0.000	0.056	0.000
p: 182	0.000	0.150	0.000	0.000	0.000

Locus: **Lsou29**

	N 30	28	26	9	4
p: 312	0.000	0.036	0.000	0.056	0.000
p: 314	0.950	0.857	0.731	0.667	0.000
p: 316	0.017	0.107	0.000	0.167	0.000
p: 318	0.017	0.000	0.000	0.111	0.000
p: 320	0.000	0.000	0.212	0.000	0.625
p: 322	0.000	0.000	0.019	0.000	0.375
p: 324	0.017	0.000	0.038	0.000	0.000

Locus: **BL12B**

N	30	31	26	7	4
p: 149	0.000	0.048	0.000	0.000	0.000
p: 151	1.000	0.952	1.000	1.000	1.000

Locus: **Lsou34**

N	30	31	24	8	4
p: 230	0.450	0.016	0.083	0.125	0.000
p: 232	0.200	0.032	0.146	0.313	0.000
p: 234	0.017	0.016	0.021	0.000	0.000
p: 236	0.100	0.661	0.479	0.313	0.000
p: 240	0.183	0.226	0.000	0.000	0.625
p: 242	0.000	0.000	0.000	0.000	0.250
p: 246	0.000	0.000	0.000	0.063	0.125
p: 248	0.033	0.016	0.000	0.188	0.000
p: 250	0.000	0.000	0.188	0.000	0.000
p: 252	0.000	0.032	0.021	0.000	0.000
p: 254	0.017	0.000	0.042	0.000	0.000
p: 256	0.000	0.000	0.021	0.000	0.000

Locus: **Lsou08**

N	28	31	25	6	4
p: 191	0.089	0.081	0.000	0.000	0.000
p: 193	0.018	0.081	0.000	0.000	0.000
p: 195	0.054	0.129	0.060	0.333	0.000
p: 197	0.250	0.323	0.520	0.500	0.750
p: 199	0.089	0.194	0.300	0.083	0.250
p: 201	0.089	0.194	0.080	0.083	0.000
p: 203	0.089	0.000	0.020	0.000	0.000
p: 207	0.286	0.000	0.020	0.000	0.000
p: 209	0.036	0.000	0.000	0.000	0.000

Locus: **LceC1**

N	30	30	25	7	4
p: 87	0.033	0.083	0.020	0.071	0.000
p: 89	0.317	0.117	0.820	0.000	0.000
p: 93	0.333	0.250	0.060	0.571	0.500
p: 95	0.017	0.133	0.040	0.214	0.000
p: 97	0.017	0.067	0.060	0.000	0.000
p: 99	0.150	0.000	0.000	0.000	0.000
p: 101	0.117	0.033	0.000	0.071	0.500
p: 103	0.017	0.317	0.000	0.071	0.000

Locus: **RSer10**

N	26	26	24	7	4
p: 183	0.077	0.000	0.042	0.000	0.000
p: 185	0.250	0.077	0.854	0.000	1.000
p: 187	0.038	0.000	0.000	0.071	0.000
p: 189	0.096	0.423	0.042	0.500	0.000
p: 191	0.038	0.481	0.000	0.143	0.000
p: 193	0.442	0.000	0.042	0.071	0.000
p: 197	0.038	0.000	0.000	0.143	0.000
p: 199	0.000	0.019	0.000	0.000	0.000
p: 201	0.019	0.000	0.000	0.000	0.000
p: 205	0.000	0.000	0.021	0.071	0.000

Locus: **N7K4**

N	30	28	26	7	4
p: 150	0.000	0.000	0.019	0.000	0.000
p: 152	0.183	0.054	0.058	0.000	0.000
p: 154	0.717	0.179	0.596	0.643	0.000
p: 156	0.033	0.000	0.192	0.071	0.375
p: 158	0.033	0.768	0.058	0.143	0.625
p: 160	0.000	0.000	0.058	0.000	0.000
p: 162	0.033	0.000	0.019	0.143	0.000

Locus**: CypG24**

N	30	30	26	8	4
p: 133	0.083	0.450	0.077	0.313	0.000
p: 137	0.017	0.000	0.058	0.000	0.000
p: 139	0.500	0.333	0.827	0.500	1.000
p: 141	0.383	0.217	0.038	0.188	0.000
p: 145	0.017	0.000	0.000	0.000	0.000

Příloha č. 3 *Frekvence genotypů v jednotlivých populacích vygenerované programem GenePop, dvě řady čísel představují určitou kombinaci alel, např. 79/79, 79/85, 85/85 atd.*

Locus: **BL130**

Pop	Genotypes:											
	79	79	85	79	85	87	87	89	85	87	89	
	79	85	85	87	87	87	89	89	93	93	93	Total
Zalomka	4	2	1	12	1	6	1	0	1	0	0	28
Cernica	0	0	1	1	4	20	1	0	0	2	1	30
Vrijeka	0	0	22	0	1	1	1	0	0	0	0	25
Mušnica	0	0	1	0	1	4	1	1	0	0	0	8
Ljuta.	0	0	4	0	0	0	0	0	0	0	0	4
Total:	4	2	29	13	7	31	4	1	1	2	1	95

Locus: **BL144**

Pop	Genotypes:					
	122	122	122	126	130	
	122	126	130	130	130	Total
Zalomka	28	0	0	0	0	28
Cernica	15	2	7	1	3	28
Vrijeka	19	0	6	0	0	25
Mušnica	7	0	0	0	0	7
Ljuta	4	0	0	0	0	4
Total:	73	2	13	1	3	92

Locus: **Lsou05**

Pop	Genotypes:																							
	177	185	185	187	189	185	187	189	191	187	191	193	193	195	185	189	191	201	195	203	185	191	207	
	177	185	187	187	189	191	191	191	191	193	193	193	195	195	201	201	201	201	203	203	205	205	207	Total
Zalomka	0	10	0	1	0	8	0	0	4	0	0	0	0	0	1	0	1	0	0	1	1	1	0	28
Cernica	0	0	0	0	0	0	0	0	28	0	1	0	0	0	0	0	0	0	0	0	0	0	0	29
Vrijeka	1	0	4	11	1	0	1	0	2	1	0	1	0	0	0	1	0	1	0	0	0	0	0	24
Mušnica	0	0	0	2	1	0	1	1	1	0	0	0	0	0	0	0	0	0	0	1	0	1	1	9
Ljuta	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	2	0	0	0	0	4
Total:	1	10	4	14	2	8	2	1	35	1	1	1	1	1	1	1	1	1	2	2	1	2	1	94

Locus: **BL161**

Pop	Genotypes: 195 197	197 197	197 199	Total
Zalomka	0	29	1	30
Cernica	1	29	0	30
Vrijeka	0	26	0	26
Mušnica	0	9	0	9
Ljutaa	0	4	0	4
Total:	1	97	1	99

Locus: **BL1153**

Pop	Genotypes: 220 220	220 222	222 222	Total
Zalomka	16	11	2	29
Cernica	21	6	1	28
Vrijeka	26	0	0	26
Mušnica	6	3	0	9
Ljuta.	4	0	0	4
Total:	73	20	3	96

Locus: **Lsou19**

Pop	Genotypes: 178 178	178 180	180 180	178 182	182 182	Total
Zalomka	15	11	4	0	0	30
Cernica	22	1	0	5	2	30
Vrijeka	26	0	0	0	0	26
Mušnica	8	1	0	0	0	9
Ljuta	4	0	0	0	0	4
Total:	75	13	4	5	2	99

Locus: **Lsou29**

Pop Genotypes:

Pop												Total
	312	312	314	314	314	318	314	320	320	314	322	
	312	314	314	316	318	318	320	320	322	324	324	Total
Zalomka	0	0	27	1	1	0	0	0	0	1	0	30
Cernica	1	0	21	6	0	0	0	0	0	0	0	28
Vrijeka	0	0	16	0	0	0	5	3	0	1	1	26
Mušnica	0	1	4	3	0	1	0	0	0	0	0	9
Ljuta	0	0	0	0	0	0	0	1	3	0	0	4
Total:	1	1	68	10	1	1	5	4	3	2	1	97

Locus: BL12B

Pop Genotypes:

Pop				Total
	149	149	151	
	149	151	151	Total
Zalomka	0	0	30	30
Cernica	1	1	29	31
Vrijeka	0	0	26	26
Mušnica	0	0	7	7
Ljuta	0	0	4	4
Total:	1	1	96	98

Locus: **Lsou34**

Pop Genotypes:

Pop																											Total
	230	230	232	230	230	232	234	236	230	232	236	240	240	240	230	240	246	248	232	236	250	232	252	236	250	232	
	230	232	232	234	236	236	236	236	240	240	240	240	242	246	248	248	248	248	250	250	250	252	252	254	254	256	Total
Zalomka	7	4	1	1	0	3	0	1	6	3	0	1	0	0	2	0	0	0	0	0	0	0	0	1	0	0	30
Cernica	0	0	1	0	0	0	1	15	1	0	10	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0	31
Vrijeka	1	0	1	1	1	1	0	8	0	0	0	0	0	0	0	0	0	0	2	4	1	1	0	1	1	1	24
Mušnica	0	0	2	0	2	1	0	1	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0	0	0	0	8
Ljuta	0	0	0	0	0	0	0	0	0	0	0	1	2	1	0	0	0	0	0	0	0	0	0	0	0	0	4
Total:	8	4	5	2	3	5	1	25	7	3	10	3	2	1	2	1	1	1	2	4	1	1	1	2	1	1	97

Locus: **Lsou08**

Pop Genotypes:

Pop	191	193	193	195	191	193	195	197	191	195	197	199	193	197	199	201	197	201	203	191	197	199	201	203	207	207	
	191	193	195	195	197	197	197	197	199	199	199	199	201	201	201	201	203	203	203	207	207	207	207	207	207	209	Total
Zalomka	0	0	0	0	2	1	3	2	0	0	1	1	0	1	1	1	1	0	1	3	1	1	1	2	3	2	28
Cernica	2	1	2	0	0	0	5	3	1	1	0	5	1	9	0	1	0	0	0	0	0	0	0	0	0	0	31
Vrijeka	0	0	0	0	0	0	3	10	0	0	2	6	0	0	1	1	0	1	0	0	1	0	0	0	0	0	25
Mušnica	0	0	0	1	0	0	2	1	0	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	6
Ljuta	0	0	0	0	0	0	0	2	0	0	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	4
Total:	2	1	2	1	2	1	13	18	1	1	6	12	1	11	2	3	1	1	1	3	2	1	1	2	3	2	94

Locus: **LceC1**

Pop Genotypes:

Pop	87	87	89	87	93	89	93	95	89	93	95	93	99	93	97	99	101	93	95	97	101	103	
	87	89	89	93	93	95	95	95	97	97	97	99	99	101	101	101	101	103	103	103	103	103	Total
Zalomka	0	2	8	0	6	1	0	0	0	0	0	5	1	3	1	2	0	0	0	0	1	0	30
Cernica	1	3	2	0	4	0	2	1	0	2	1	0	0	0	0	0	0	3	3	1	2	5	30
Vrijeka	0	1	18	0	1	2	0	0	2	1	0	0	0	0	0	0	0	0	0	0	0	0	25
Mušnica	0	0	0	1	2	0	1	1	0	0	0	0	0	1	0	0	0	1	0	0	0	0	7
Ljuta	0	0	0	0	1	0	0	0	0	0	0	0	0	2	0	0	1	0	0	0	0	0	4
Total:	1	6	28	1	14	3	3	2	2	3	1	5	1	6	1	2	1	4	3	1	3	5	96

Locus: **RSer10**

Pop Genotypes:

Pop	183	185	187	185	187	189	189	191	183	185	189	191	193	189	193	191	191	189	197	
	185	185	187	189	189	189	191	191	193	193	193	193	193	197	197	199	201	205	205	Total
Zalomka	1	5	1	1	0	0	0	0	3	1	4	1	6	0	2	0	1	0	0	26
Cernica	0	2	0	0	0	6	10	7	0	0	0	0	0	0	0	1	0	0	0	26
Vrijeka	2	19	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	24
Mušnica	0	0	0	0	1	1	2	0	0	0	1	0	0	1	0	0	0	0	1	7
Ljuta	0	4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	4
Total:	3	30	1	2	1	7	12	7	3	1	5	1	7	1	2	1	1	1	1	87

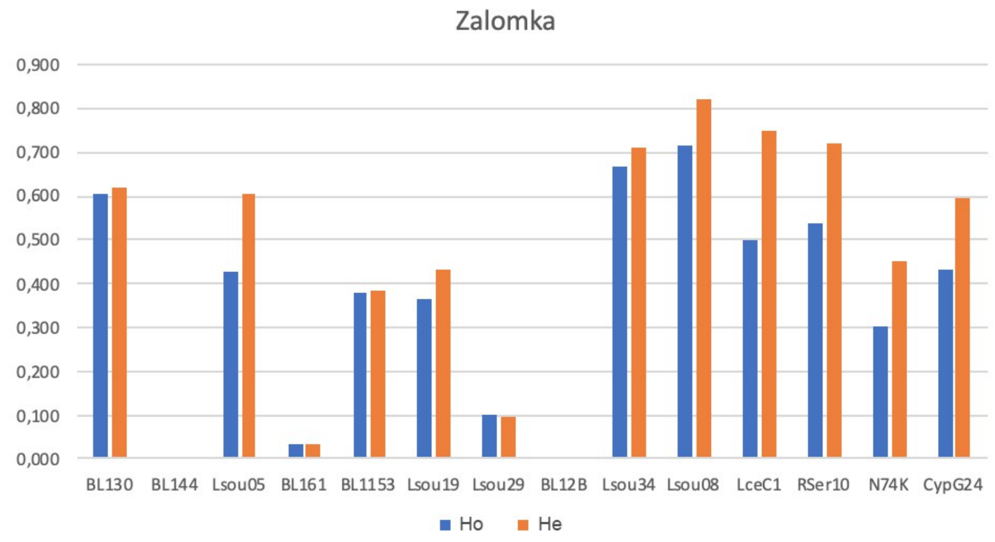
Locus: **N7K4**

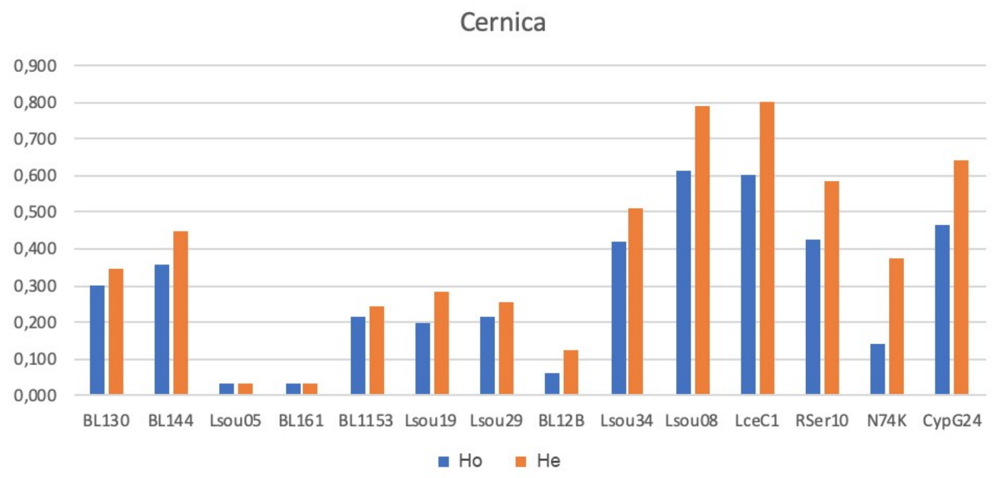
Pop	Genotypes:													
	152	154	154	156	152	154	156	158	154	156	154			
	152	154	154	154	156	156	158	158	158	158	160	160	162	Total
Zalomka	3	0	5	17	0	1	0	2	0	0	0	0	2	30
Cernica	1	0	1	3	0	0	0	3	0	20	0	0	0	28
Vrijeka	0	1	2	9	6	1	1	2	0	0	1	2	1	26
Mušnica	0	0	0	3	1	0	0	0	0	1	0	0	2	7
Ljuta	0	0	0	0	0	0	0	0	3	1	0	0	0	4
Total:	4	1	8	32	7	2	1	7	3	22	1	2	5	95

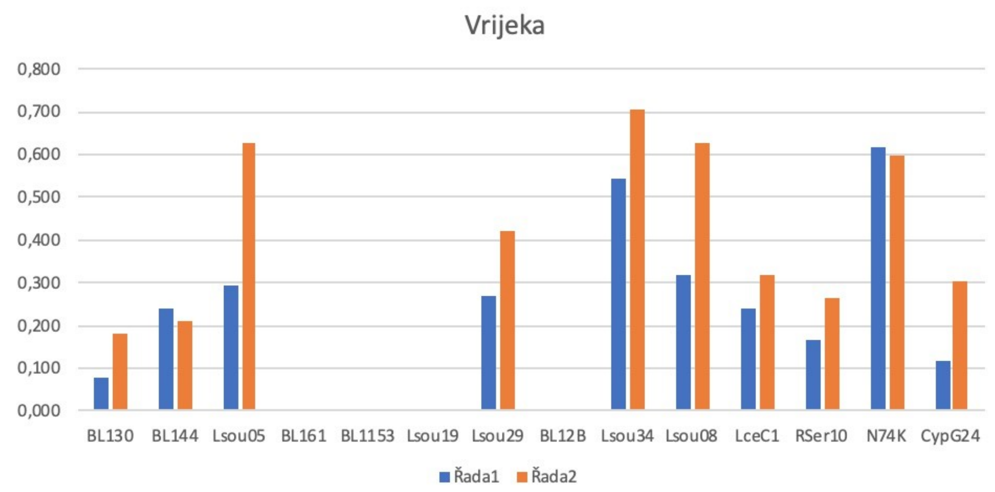
Locus: **CypG24**

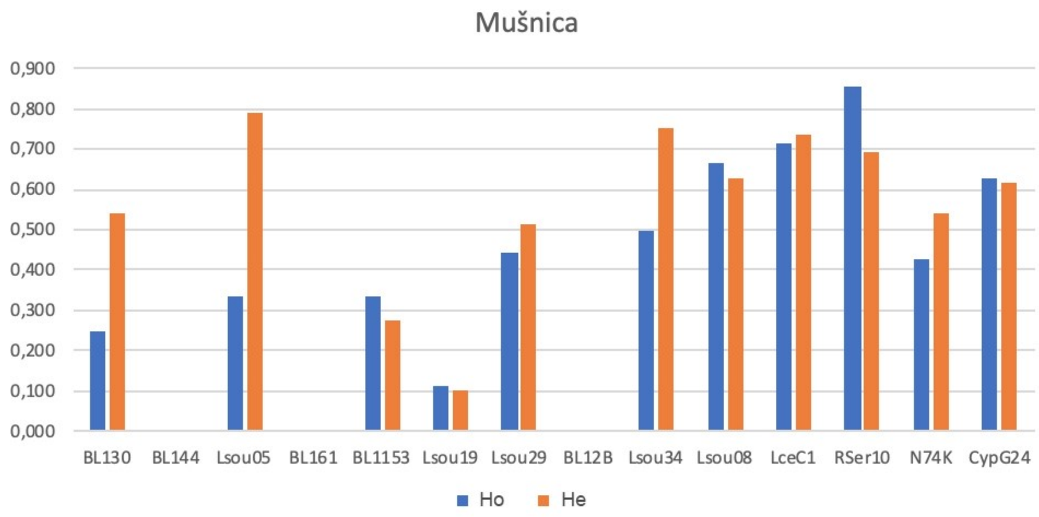
Pop	Genotypes:									
	133	133	133	137	139	133	139	141	139	
	133	137	139	139	139	141	141	141	145	Total
Zalomka	0	1	4	0	9	0	7	8	1	30
Cernica	8	0	5	0	6	6	3	2	0	30
Vrijeka	2	0	0	3	20	0	0	1	0	26
Mušnica	1	0	2	0	2	1	2	0	0	8
Ljuta	0	0	0	0	4	0	0	0	0	4
Total:	11	1	11	3	41	7	12	11	1	98

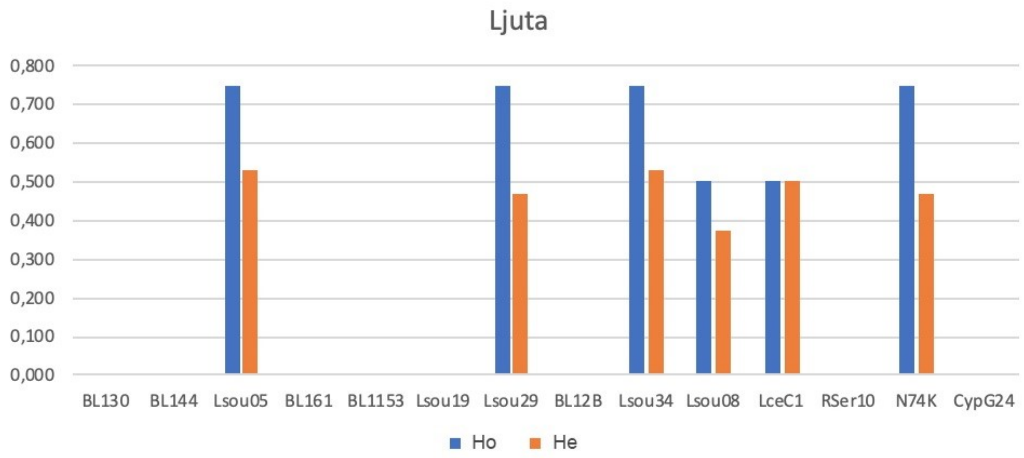
Příloha č. 4 *Grafy znázorňující pozorovanou a očekávanou heterozygotnost v lokusech jednotlivých populací spočítané v programu GenAlEx a zpracované v programu Microsoft Excel. Ho = pozorovaná, He = očekávaná.*








Mušnica
0,900
0,800
0,700
0,600
0,500
0,400
0,300
0,200
0,100
0,000
BL130 BL144 Lsou05 BL161 BL1153 Lsou19 Lsou29 BL12B Lsou34 Lsou08 LceC1 RSer10 N74K CypG24
Ho He


Ljuta
0,800
0,700
0,600
0,500
0,400
0,300
0,200
0,100
0,000
BL130 BL144 Lsou05 BL161 BL1153 Lsou19 Lsou29 BL12B Lsou34 Lsou08 LceC1 RSer10 N74K CypG24
Ho He